

Purine Nucleosides. XXIX. The Synthesis of 2'-Deoxy-L-adenosine and 2'-Deoxy-L-guanosine and Their α Anomers^{1a}

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The synthesis of 6-amino-9-(2-deoxy- β -L-erythro-pentofuranosyl)purine (2'-deoxy-L-adenosine) (9) and its α anomer 8 has been accomplished by the first reported fusion of a 1-O-methyl-2-deoxy sugar derivative. Fusion of 1-O-methyl-3,5-di-O-p-toluy-2-deoxy-L-erythro-pentofuranose (1) and 2,6-dichloropurine (2) gave 2,6-dichloro-9-(3,5-di-O-p-toluy-2-deoxy- α - and - β -L-erythro-pentofuranosyl)purines (3 and 4, respectively). Selective amination at position 6 with concurrent deblocking, followed by hydrogenolysis of the 2-chloro function, gave the desired L enantiomers 8 and 9. The α and β anomers of 1 were separated and individually fused with 2. The α anomer gave higher total yields of nucleosides (3 plus 4) and gave a higher proportion of β nucleoside 4. Fusion of 1-O-acetyl-3,5-di-O-p-toluy-2-deoxy-L-erythro-pentofuranose (5) and 2-fluoro-6-benzyloxy-purine (10) followed by treatment with alcoholic ammonia and hydrogenolysis of the 6-benzyloxy group gave 2-amino-9-(2-deoxy- β -L-erythro-pentofuranosyl)purin-6-one (2'-deoxy-L-guanosine, 15) and its α anomer 12. These 2' deoxynucleosides obey Hudson's isorotation rule and the "triplet"- "quartet" ¹H nmr anomeric proton splitting patterns for β and α anomers, respectively.

The synthesis of L-adenosine² and DL-adenosine³ represent the first attempts to prepare ribonucleosides for biological and physical investigation of enantiomeric nucleic acid components. During the course of this work,⁴ a report of the preparation of L-thymidine appeared.⁵

We now wish to report the synthesis of 2'-deoxy- α - and - β -L-adenosines and -guanosines, which are the first examples of enantiomorphs of the natural purine deoxynucleosides of DNA. The polymerization of the β anomers of these L isomers into DNA-like fragments would provide exciting information⁶ concerning helical structure and properties. The finding that 6-amino-9-(2-deoxy- α -L-erythro-pentofuranosyl)purine (8) acts as a substrate for adenosine deaminase⁷ suggests the potential biological activity of stereoisomers of deoxynucleosides, a possibility borne out in the case of the selectively toxic 2-amino-9-(2-deoxy- α -D-erythro-pentofuranosyl)purine-6-thione (2'-deoxy- α -thioguanosine).⁸

Success of the fusion⁹ procedure for purine deoxynucleoside synthesis^{10,11} suggested application of this method to the 2'-deoxy-L isomers. The preparation of 2-deoxy-L-erythro-pentose (2-deoxy-L-ribose) was effected according to the procedure of Vargha and Kuszman¹² (for the D enantiomer) from 3,5-di-O-acetyl-L-arabinal.¹³ The method of Hoffer¹⁴ was used to

convert the free 2-deoxy-L-erythro-pentose into 1-O-methyl-3,5-di-O-p-toluy-2-deoxy-L-erythro-pentofuranose (1).

The 1-O-methyl sugar 1 (Scheme I) was fused directly with 2,6-dichloropurine (2) to give the anomeric 2,6-dichloro-9-(3,5-di-O-p-toluy-2-deoxy- α - and - β -L-erythro-pentofuranosyl)purines (3 and 4, respectively), which were resolved into pure anomers by alumina column chromatography and fractional crystallization. Treatment of these anomeric nucleosides with alcoholic ammonia gave 6-amino-2-chloro-9-(2-deoxy- α - and - β -L-erythro-pentofuranosyl)purines (6 and 7, respectively). These anomerically pure intermediates were catalytically hydrogenated to give 6-amino-9-(2-deoxy- β -L-erythro-pentofuranosyl)purine (2'-deoxy-L-adenosine, 9) and 6-amino-9-(2-deoxy- α -L-erythro-pentofuranosyl)purine (8).

This sequence represents the first reported use of a 1-O-methyl-2-deoxy sugar derivative in the fusion synthesis of deoxynucleosides. The α and β anomers of 1 were resolved by fractional crystallization and were individually subjected to fusion with 2,6-dichloropurine (2). Dichloroacetic acid catalyzed fusion of 1 (β anomer) and 2 at 140° for 5 min gave a 15% isolated yield of the blocked nucleosides 3 (63%) and 4 (37%). Identical fusion of 1 (α anomer) and 2 gave a 45% isolated yield of 3 (29%) and 4 (71%). Similar fusion of the anomeric mixture of 1 with 2 gave a 25% yield of 3 (41%) and 4 (59%). These results indicate that 1-O-methyl-3,5-di-O-p-toluy-2-deoxy- α -L-erythro-pentofuranose is structurally more suitable for the fusion reaction and also leads to the predominant formation of β nucleoside 4 by overall inversion at C-1.

The α - and β -L-2'-deoxyadenosines (8 and 9, respectively) were found to exhibit identical uv, ir, and ¹H nmr spectra with their corresponding D enantiomers^{10,15} and essentially equal and opposite optical rotations (and circular dichroism spectra¹⁶).

For the synthesis of the anomeric L-2'-deoxyguanosines, a recently developed method for guanine nucleoside synthesis¹¹ was employed. The anomeric mixture of 1 was hydrolyzed with dilute acid to give 3,5-di-O-p-toluy-2-deoxy-L-erythro-pentose, which was acetylated

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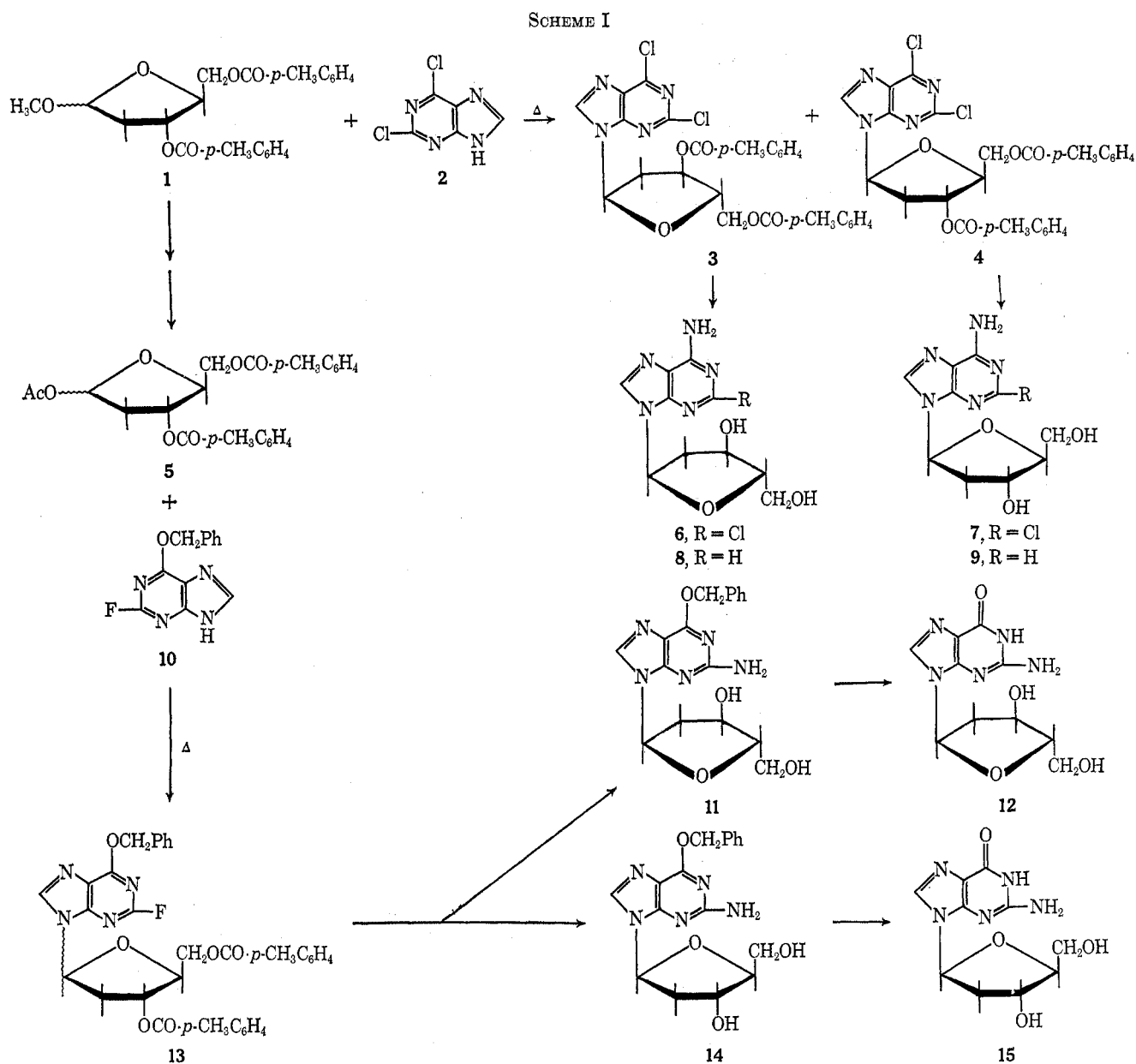
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to give 1-*O*-acetyl-3,5-di-*O*-*p*-toluyloxy-2-deoxy-*L*-erythro-pentofuranose⁵ (**5**) as a sirupy mixture. Fusion of **5** (ca. 50:50 α/β by nmr) with 2-fluoro-6-benzyloxy-purine¹¹ (**10**) gave at least a 14% yield of the anomeric nucleoside **13**. This intermediate was verified by uv and tlc and was then treated directly with alcoholic ammonia. The resulting 2-amino-6-benzyloxy-9-(2-deoxy- α - and - β -*L*-erythro-pentofuranosyl)purines (**11** and **14**, respectively) were resolved by chromatography on Dowex 1-X2 (OH⁻).^{11,17} The observed ratio of α/β anomers in this case was ca. 2:1, which is in contrast with the predominance of β anomer in the fusion of **1** and **2**. As in the case of the *D* enantiomers,¹¹ **11** (α anomer) crystallized and was completely characterized. Hydrogenation of the anomericly pure intermediates **14** and **11** over palladium gave 2-amino-9-(2-deoxy- β -*L*-erythro-pentofuranosyl)purin-6-one (2'-deoxy-*L*-guanosine, **15**) and 2-amino-9-(2-deoxy- α -*L*-erythro-pentofuranosyl)purin-6-one (**12**).

Again these products were characterized and their structures were confirmed by comparison with the

corresponding *D* enantiomers.¹¹ It is of interest to note that (as expected) the H_1' proton of these *L*-2'-deoxynucleosides obey the same "triplet"- "quartet" splitting patterns for β and α anomers, respectively, as observed with a number of previously observed^{10,11} *D*-2'-deoxynucleosides.

Experimental Section

Melting points were determined on a Fisher-Johns block and are uncorrected. Nmr spectra were determined on a Varian A-60 instrument with tetramethylsilane or sodium 5,5-dimethyl-5-silapentanesulfonate as internal standard. Uv spectra were determined on a Beckman DK-2 instrument. Hydrogenations were effected using a Parr hydrogenation apparatus at specified hydrogen gas pressure. Evaporations were accomplished using a Büchler rotating evaporator under reduced pressure (aspirator) unless specified otherwise. Thin layer chromatography (tlc) was run on glass plates coated with SilicAR-7GF (Mallinckrodt Chemical Works) using the upper phase of EtOAc-*n*-PrOH-H₂O (4:1:2) unless otherwise specified.

1-*O*-Methyl-3,5-di-*O*-*p*-toluyloxy-2-deoxy-*L*-erythro-pentofuranose (1).—To 280 ml of H₂O was added 10 g (0.048 mol) of 2-deoxy-*L*-erythro-pentose anilide,⁵ 10 ml of benzaldehyde, and 1 g of benzoic acid. This mixture was stirred for 17 hr at room temperature and then extracted with three 100-ml portions of Et₂O. The resulting aqueous solution was evaporated to dryness at a temperature less

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than 30° and EtOH was added to the residue. This solution was evaporated to dryness and this procedure was repeated twice with absolute EtOH and twice with absolute MeOH. The resulting 2-deoxy-*L*-erythro-pentose was dissolved in 100 ml of absolute MeOH and treated with MeOH-HCl followed by *p*-toluyl chloride according to the procedure of Hoffer.¹⁴ The resulting 1-*O*-methyl-3,5-di-*O*-*p*-toluyl-2-deoxy-*L*-erythro-pentofuranose (1) was dissolved in 20 ml of absolute EtOH and cooled at 0°. Three crops of crystalline sugar (5.2 g, 4.2 g, and 2.9 g, respectively, total yield 67%) were obtained. Several recrystallizations of the first crop from EtOH gave colorless needles of 1-*O*-methyl-3,5-di-*O*-*p*-toluyl-2-deoxy- α -*L*-erythro-pentofuranose (α anomer of 1): mp 83–84°; $[\alpha]^{25}_D -135.2^\circ$ (*c* 0.9, CHCl₃); nmr (CDCl₃) δ 3.40 (s, 3, OCH₃-1) [lit.¹⁵ (for the *D* enantiomer) mp 82–83°; $[\alpha]^{20}_D +130^\circ$ (*c* 0.67, CHCl₃)].

Anal. Calcd for C₂₂H₂₄O₆: C, 68.76; H, 6.25. Found: C, 68.48; H, 6.21.

Several recrystallizations of the third crop of crystalline 1 gave needles of 1-*O*-methyl-3,5-di-*O*-*p*-toluyl-2-deoxy- β -*L*-erythro-pentofuranose (β anomer of 1): mp 78–78.5°; $[\alpha]^{25}_D +7.7^\circ$ (*c* 1.7, CHCl₃); nmr (CDCl₃) δ 3.32 (s, 3, OCH₃-1) [lit.¹⁵ (for the *D* enantiomer) mp 76.5–78°; $[\alpha]^{20}_D -8.1^\circ$ (*c* 2.5, CHCl₃)].

Anal. Found: C, 68.95; H, 6.37.

2,6-Dichloro-9-(3,5-di-*O*-*p*-toluyl-2-deoxy- α - and - β -*L*-erythro-pentofuranosyl)purines (3 and 4). A. From 1-*O*-Methyl-3,5-di-*O*-*p*-toluyl-2-deoxy- α -*L*-erythro-pentofuranose (α Anomer of 1).—A finely powdered mixture of 1.87 g (0.0049 mol) of 1-*O*-methyl-3,5-di-*O*-*p*-toluyl-2-deoxy- α -*L*-erythro-pentofuranose and 0.93 g (0.0049 mol) of 2,6-dichloropurine (2) was heated in an oil bath at 143° for 6 min. Dichloroacetic acid (2 drops) was added and fusion was continued for 5 min at 143° *in vacuo* (aspirator). The clear melt was cooled to about 100° and dissolved in EtOAc. This solution was washed with two 50-ml portions of cold, saturated, aqueous NaHCO₃ and 50 ml of cold H₂O, dried (Na₂SO₄), and filtered. The filtrate was evaporated to a gum and this material was treated twice with EtOH and evaporated. The residue was dissolved in 5 ml of benzene and this solution was applied to a neutral alumina column (90 g). The column was washed with 1000 ml of benzene and elution was begun with EtOAc-PhH (2:8). The fractions (100 ml) were evaporated to dryness and evaluated by uv and tlc. Fractions 1 and 2 contained 0.33 g of sugar 1; fractions 3–9 contained the blocked nucleosides and were fractionally recrystallized individually from EtOH to give 0.35 g (13%) of 3 (α anomer) and 0.84 g (32%) of 4 (β anomer), total yield 1.19 g (45%). Pure 2,6-dichloro-9-(3,5-di-*O*-*p*-toluyl-2-deoxy- α -*L*-erythro-pentofuranosyl)purine (3) was obtained, mp 140–142°, uv max (EtOH) 241 m μ (ϵ 35,800) and 272.5 (11,500).

Anal. Calcd for C₂₈H₂₂O₅N₄Cl₂: C, 57.68; H, 4.06; N, 10.35. Found: C, 57.72; H, 4.06; N, 10.37.

Pure 2,6-dichloro-9-(3,5-di-*O*-*p*-toluyl-2-deoxy- β -*L*-erythro-pentofuranosyl)purine (4) was obtained, mp 154–156°, uv max (EtOH) 240.5 m μ (ϵ 35,500) and 272.5 (11,300).

Anal. Found: C, 57.55; H, 4.21; N, 10.45.

B. From 1-*O*-Methyl-3,5-di-*O*-*p*-toluyl-2-deoxy- β -*L*-erythro-pentofuranose (β Anomer of 1).—Fusion of 1.67 g (0.00435 mol) of 1-*O*-methyl-3,5-di-*O*-*p*-toluyl-2-deoxy- β -*L*-erythro-pentofuranose and 0.83 g (0.0044 mol) of 2,6-dichloropurine (2) according to procedure A above gave 0.22 g (9.3%) of 3 (α anomer) and 0.13 g (5.5%) of 4 (β anomer), total yield 14.8%.

C. From 1-*O*-Methyl-3,5-di-*O*-*p*-toluyl-2-deoxy-*L*-erythro-pentofuranose (1).—Fusion of 2.7 g (0.007 mol) of the crystalline anomeric mixture 1 with 1.3 g (0.0069 mol) of 2,6-dichloropurine (2) according to procedure A above gave 0.38 g (10%) of 3 (α anomer) and 0.55 g (15%) of 4 (β anomer), total yield 25%.

6-Amino-9-(2-deoxy- α -*L*-erythro-pentofuranosyl)purine (8).—To a solution of 100 ml of methanol saturated with ammonia at room temperature was added 1.47 g (0.0027 mol) of 3 and the suspension was stirred at room temperature for 3 days with periodic addition of ammonia gas to saturation. The resulting solution was heated on the steam bath for 30 min and then evaporated to dryness. The residue was treated with 100 ml of H₂O and this was washed with three 100-ml portions of Et₂O. The aqueous solution of 6-amino-2-chloro-9-(2-deoxy- α -*L*-erythro-pentofuranosyl)purine (6) had uv absorption and tlc mobility identical with those of the corresponding *D* enantiomer¹⁰ and was hydrogenated without further purification.

The above aqueous solution was diluted to 150 ml with H₂O and 15 ml of concentrated, aqueous NH₃ was added. The resulting solution was hydrogenated at 40 psi for 8 hr in the presence of 1 g of 10% Pd-C. This mixture was filtered and the filtrate was evaporated with a water bath at less than 25°. The residue was dissolved in 1.5 ml of H₂O, cooled at 0° for 16 hr, and filtered to give 0.21 g (31%) of 8. A second crop, 0.11 g, raised the yield to 47%. Recrystallization of this material from H₂O gave 8 as needles: mp 204–204.5°; $[\alpha]^{25}_D -70.8^\circ$ (*c* 1.0, H₂O) [lit.¹⁵ (for the *D* enantiomer) $[\alpha]^{25}_D +68.2^\circ$ (H₂O)]; uv max (pH 1) 257 m μ (ϵ 16,000), (pH 11) 259 m μ (ϵ 16,400); nmr (D₂O) δ 6.42 ("q," 1, $J_{1'-2',2''} = 3.3$ and 7.5 Hz, H_{1'}).

Anal. Calcd for C₁₀H₁₃N₅O₅: C, 47.80; H, 5.22; N, 27.88. Found: C, 47.91; H, 5.38; N, 27.96.

6-Amino-9-(2-deoxy- β -*L*-erythro-pentofuranosyl)purine (2'-Deoxy-*L*-adenosine, 9).—Treatment of 1.19 g (0.0022 mol) of 4 with methanolic ammonia followed by hydrogenation under the identical conditions described above for the α anomer (3 \rightarrow 8) gave 0.21 g (38%) of pure, crystalline 9: mp 184–185°; $[\alpha]^{25}_D +23.2^\circ$ (*c* 1, H₂O) [lit.¹⁵ (for the *D* enantiomer) $[\alpha]^{25}_D -24.0^\circ$ (H₂O)]; uv max (pH 1) 257 m μ (ϵ 15,400); (pH 11) 260 m μ (ϵ 15,800); nmr (D₂O) δ 6.42 ("t," 1, $J_{1'-2',2''} = 7.0$ Hz, H_{1'}).

Anal. Calcd for C₁₀H₁₃N₅O₅: C, 47.80; H, 5.22; N, 27.88. Found: C, 47.72; H, 5.43; N, 28.00.

2-Amino-6-benzyloxy-9-(2-deoxy- α -*L*-erythro-pentofuranosyl)purine (11) and 2-Amino-6-benzyloxy-9-(2-deoxy- β -*L*-erythro-pentofuranosyl)purine (14).—A well-stirred mixture of 4.22 g (0.017 mol) of finely powdered 2-fluoro-6-benzyloxypurine¹¹ (10) and 7.82 g (0.019 mol) of sirupy 1-*O*-acetyl-3,5-di-*O*-*p*-toluyl-2-deoxy-*L*-erythro-pentofuranose⁵ (5) was placed in an oil bath preheated to 155°. Dichloroacetic acid (7 drops) was added with stirring and the mixture was stirred for 8 min at 155°, at which time a clear, amber melt had formed. An aspirator was connected and fusion *in vacuo* was continued for 17 min. The melt was cooled to ca. 100° and dissolved in EtOAc. This solution was washed with two 50-ml portions of ice-cold, saturated, aqueous Na₂CO₃ solution, 50 ml of ice-H₂O, and 50 ml of saturated aqueous NaCl solution, and dried (Na₂SO₄). This mixture was filtered using a Norit-Celite bed and the filtrate was evaporated to a heavy sirup. MeOH was added and evaporated and this was repeated twice. The sirup was dissolved in 25 ml of MeOH, treated with 200 ml of MeOH presaturated with NH₃ at -10°, and heated at 85° for 4 hr in a steel bomb. The solution was cooled, 17 ml of 1 *N* NaOH was added, and the solution was evaporated to dryness. The residue was treated with 100 ml of EtOAc and 40 ml of H₂O and the separated aqueous layer was extracted with three 50-ml portions of EtOAc. The combined organic phase was washed with 30 ml of saturated aqueous NaCl, dried (Na₂SO₄), filtered, and evaporated to ca. 20 ml. This solution was applied to a column (2 \times 20 in.) of silica gel, the column was washed with 800 ml of CHCl₃ to remove *p*-toluamide and methyl *p*-toluate, and the nucleoside material was eluted with EtOH. The EtOH fractions were evaporated to dryness, the residue was dissolved in 9 ml of 1,2-dimethoxyethane (glyme), and 11 ml of H₂O was added. This solution was applied to a column (1 \times 35 in., 500 ml) of Dowex 1-X2 (OH⁻) 200–400 mesh¹⁷ packed in glyme-H₂O (45:55).¹¹ Elution was effected with the same solvent mixture and 10-ml fractions were collected. Fractions 1–72 were discarded. Fractions 73–89 were pooled and evaporated to dryness to yield crude 2-amino-6-benzyloxy-9-(2-deoxy- α -*L*-erythro-pentofuranosyl)purine (11). This material was recrystallized from *i*-PrOH using seed crystals of the *D* enantiomer to give 0.1 g (1.6% overall yield from 10) of fine needle clusters: mp 97–99° [lit.¹¹ (for the *D* enantiomer) mp 158–160°]; uv max (pH 1) 287 m μ (ϵ 12,500), (pH 11) 280 m μ (ϵ 12,000) and 249 (10,000), (MeOH) 282 m μ (ϵ 12,500) and 249 (10,900); nmr (DMSO-*d*₆) δ 6.23 ("q," 1, $J_{1'-2',2''} = 3.0$ and 7.5 Hz, H_{1'}).

Anal. Calcd for C₁₇H₁₉N₅O₄: C, 57.13; H, 5.36; N, 19.60. Found: C, 57.33; H, 5.21; N, 19.59.

Fractions 90–97 contained both anomers and were discarded.

Fractions 98–130 were pooled and evaporated to dryness to give crude 2-amino-6-benzyloxy-9-(2-deoxy- β -*L*-erythro-pentofuranosyl)purine (14), which was hydrogenated without further purification. The tlc migrations of 11 and 14 were identical with those of their *D* enantiomers,¹¹ $R_{14}/R_{11} = 1.1$.

2-Amino-9-(2-deoxy- α -*L*-erythro-pentofuranosyl)purin-6-one (12).—The combined filtrates from crystallization of 11 were evaporated to dryness, dissolved in 25 ml of EtOH and 50 ml of H₂O, and hydrogenated for 17 hr at 46 psi with 0.12 g of 5%

(18) D. L. MacDonald and H. G. Fletcher, Jr., *J. Amer. Chem. Soc.*, **84**, 1262 (1962).

Pd-C. The mixture was filtered using Celite and the filtrate was evaporated to dryness. The white crystalline solid was recrystallized from 6 ml of H₂O to give 0.36 g (7.3% overall yield from 10) of 12 hemihydrate: $[\alpha]^{25}_D -103^\circ$ (c 1.1, DMF) [lit.¹¹ (for the D enantiomer) $[\alpha]^{25}_D +102.4^\circ$ (c 0.99, DMF)]; uv max (pH 1) 253 m μ (ϵ 12,700) and 274 sh (8800), (pH 11) 258-265 m μ (br, ϵ 12,000), (MeOH) 253 m μ (ϵ 14,500); nmr (DMSO-*d*₆-D₂O) δ 6.13 ("q," 1, $J_{1'-2',2''} = 3.5$ and 7.5 Hz, H_{1'}), (DMSO-*d*₆) δ 3.41 (s, 1, $\frac{1}{2}$ H₂O of hydration).

Anal. Calcd for C₁₀H₁₃N₅O₄· $\frac{1}{2}$ H₂O: C, 43.47; H, 5.11; N, 25.35. Found: C, 43.51; H, 4.78; N, 25.37.

2-Amino-9-(2-deoxy- β -L-erythro-pentofuranosyl)purin-6-one (2'-Deoxy-L-guanosine, 15).—The entire crude sample of 14 was dissolved in 20 ml of EtOH and 40 ml of H₂O and hydrogenated at 47 psi for 15 hr in the presence of 0.09 g of 5% Pd-C. This mixture was treated as in the preparation of 12 above to yield

0.19 g (3.9% overall yield from 10) of crystalline 15 monohydrate: $[\alpha]^{25}_D +20.5^\circ$ (c 1, DMF) [lit.¹¹ (for the D enantiomer) $[\alpha]^{25}_D -20.3^\circ$ (c 1.2, DMF)]; uv max (pH 1) 254 m μ (ϵ 12,900) and 275 sh (8900), (pH 11) 259-266 m μ (br, ϵ 12,000), (MeOH) 254 m μ (ϵ 14,700); nmr (DMSO-*d*₆-D₂O) δ 6.18 ("t," 1, $J_{1'-2',2''} = 7$ Hz, H_{1'}), nmr (DMSO-*d*₆) δ 3.46 (s, 2, H₂O of hydration).

Anal. Calcd for C₁₀H₁₃N₅O₄·H₂O: C, 42.10; H, 5.30; N, 24.55. Found: C, 41.97; H, 5.24; N, 24.53.

The anomers 12 and 15 exhibited identical tlc mobility with their D enantiomer,¹¹ $R_{15}/R_{12} = 1.2$.

Registry No.— α anomer of 1, 22837-36-1; β anomer of 1, 22837-37-2; 3, 22837-38-3; 4, 22837-39-4; 8, 17015-19-9; 9, 14365-45-8; 11, 22837-42-9; 12, 22837-43-0; 15, 22837-44-1.

The Hydrolysis of Cyclic Vinyl Ethers. An ¹⁸O Study of the Hydrolysis of 2-Alkyl-2,3,4,5,6,7-hexahydrobenzofurans¹

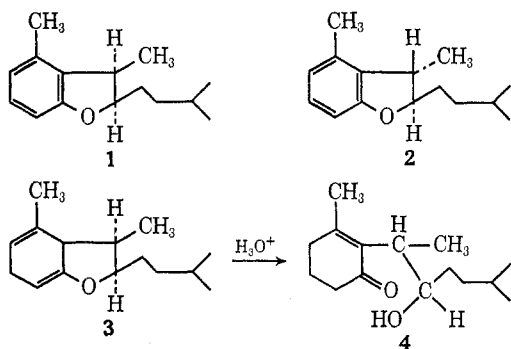
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The hydrolysis of the ¹⁸O-labeled cyclic vinyl ethers, 2-methyl-2,3,4,5,6,7-hexahydrobenzofuran (5a) and the corresponding 2,2-dimethyl compound (5b), followed by recyclization, leads to no loss of the ¹⁸O label, within experimental error of the mass spectrometric analysis. The cracking patterns for these vinyl ethers and of 2-(2'-methoxypropyl)cyclohexanone (8) have been determined. The labeling experiments rule out a free carbonium ion intermediate in the hydrolysis of 5b, where a tertiary carbonium ion could be formed; they also show that stereochemistry would be preserved around the oxygen-C-2 bond of compounds like 5a and 5b during acid hydrolysis.

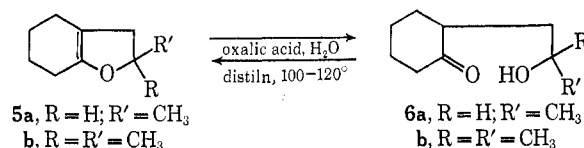
Earlier papers² have reported experiments on the preparation of 2,3-dihydrobenzofurans as possible intermediates for syntheses in the fumagillin series. One of the sequences planned involved a Birch reduction^{2c,3} of the 2,3-dialkyl-2,3-dihydrobenzofuran, such as 1, followed by hydrolysis of the resulting tetrahydrobenzofuran 3; both 1 and the corresponding *trans* compound 2 were prepared, their configurations were established, and both were reduced with lithium and liquid ammonia and then hydrolyzed.^{2,3} It is obviously



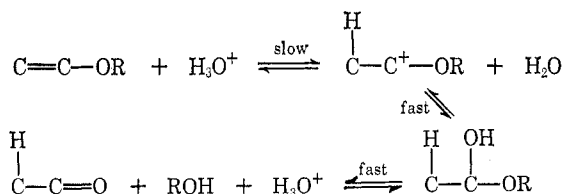
necessary to know whether in the hydrolysis of the vinyl ether 3 (and the related *trans* compound) there has been cleavage of the oxygen-C-2 bond in 3, and hence any possibility of change in the configuration of the carbon carrying the hydroxyl group in 4.

The present study shows by ¹⁸O labeling studies that there is no oxygen-C-2 cleavage in compound 5a, where

C-2 carries one alkyl group, and also none in compound 5b, where C-2 is a tertiary carbon, carrying two methyl groups.



Earlier studies on the mechanism of hydrolysis of acetals and of open-chain vinyl ethers have shown that, in H₂¹⁸O, none of the label appears in the alcohol formed,⁴ and therefore the hydrolysis does not involve cleavage of the O-R bond. Kinetic studies^{5,6} and solvent isotope⁶ effects indicate that the slow step is the transfer of a proton to the unsaturated carbon β to the oxygen atom, to form the resonance-stabilized carbo-



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